

## COURSE OUTLINE

### 1. GENERAL

<b>SCHOOL</b>	AGRICULTURAL SCIENCES		
<b>DEPARTMENT</b>	FOOD SCIENCE AND NUTRITION		
<b>EDUCATION LEVEL</b>	<i>Undergraduate</i>		
<b>LECTURE CODE</b>	ME618	<b>SEMESTER</b>	6 <sup>th</sup>
<b>LECTURE TITLE</b>	Molecular Diagnostics in Food Science		
<b>SELF-ENDED TEACHING ACTIVITIES</b>		<b>WEEKLY TEACHING HOURS</b>	<b>CREDIT UNITS (ECTS)</b>
<b>LECTURES</b>		3	3
<b>LABORATORY EXERCISES</b>		2	2
		5	5
<b>COURSE TYPE</b>	Scientific Area of Biology, Molecular Biology and Bioinformatics		
<b>PREREQUISITE COURSES:</b>	Biology, Molecular Biology and Bioinformatics		
<b>LANGUAGE OF INSTRUCTION and EXAMINATIONS:</b>	Greek		
<b>THE COURSE IS OFFERED TO ERASMUS STUDENTS</b>	Yes (English)		
<b>COURSE WEBSITE (URL)</b>	<a href="https://food.uth.gr/theodoros-goulas/">https://food.uth.gr/theodoros-goulas/</a>		

### 2. LEARNING OUTCOMES

<b>Learning outcomes</b>
<p>Upon successful completion of the course, the student will be able to:</p> <ul style="list-style-type: none"> <li>• To know the most important molecular techniques used in food and beverage safety and quality</li> <li>• To know the applications of these in different foods with detailed examples.</li> <li>• Have a good knowledge of molecular analysis, sampling, approximations and sizes.</li> <li>• To know the basic equipment for the application of molecular analysis methods.</li> </ul>
<b>General Skills</b>
<ol style="list-style-type: none"> <li>1. Search, analysis and synthesis of data and information, also using the necessary technologies.</li> <li>2. Adaptation to new situations.</li> <li>3. Decision making.</li> <li>4. Autonomous work.</li> <li>5. Group work.</li> <li>6. Generation of new research ideas.</li> <li>7. Project planning and management.</li> <li>8. Exercise criticism and self-criticism</li> <li>9. Promotion of free, creative and inductive thinking</li> </ol>

### 3. COURSE CONTENT

#### Theory

##### **1st Week**

Molecular techniques in food.

##### **2nd Week**

Unraveling the pathogenic behavior of plant pathogens through advanced molecular techniques

##### **3rd Week**

Molecular Characterization of Ochratoxigenic Fungal Flora as an Innovative Tool to Certify Coffee Origin

##### **4th Week**

Molecular and “Omics” Techniques for Studying Gut Microbiota Relevant to Food Animal Production

##### **5th Week**

Molecular identification and distribution of yeasts in fruit

##### **6th Week**

Current and new ideas on molecular methods for the identification of microbial growth in fruit juices

##### **7th Week**

Molecular techniques related to the identification of the bacterial microflora of seafood

##### **8th Week**

Assessment of the Microbial Ecology of Meat and Meat Products at the Molecular Level

##### **9th Week**

Molecular techniques for the determination of LAB in fermented cereals and meat products

##### **10th Week**

Determining the geographical origin of food with molecular techniques

##### **11th Week**

Molecular determination of enteric viruses in fresh produce

##### **12th Week**

Rapid detection of food pathogens using molecular methods 343

##### **13th Week**

Biosensor-based techniques: A reliable and primary tool for the detection of foodborne pathogens

## **Laboratory Exercises**

### **1<sup>st</sup> Week**

Real-time Reverse Transcription PCR

### **2<sup>nd</sup> Week**

Quantitative PCR

### **3<sup>rd</sup> Week**

ELISA

### **4<sup>th</sup> Week**

Fluorescence in situ hybridization (FISH)

### **5<sup>th</sup> Week**

Rapid Amplified Polymorphic DNA (RAPD)

### **6<sup>th</sup> Week**

Terminal Restriction Fragment Length Polymorphism (TRFLP)

### **7<sup>th</sup> Week**

Denaturing Gradient Gel Electrophoresis (DGGE)

### **8<sup>th</sup> Week**

Temperature Gradient Gel Electrophoresis (TGGE)

### **9<sup>th</sup> Week**

Ribosomal Intergenic Space Analysis (RISA)

### **10<sup>th</sup> Week**

Next Generation Sequencing

### **11<sup>th</sup> Week**

Microarrays

### **12<sup>th</sup> Week**

Student presentations

### **13<sup>th</sup> Week**

Student presentations and review of laboratory exercises

#### 4. TEACHING AND LEARNING METHODS - ASSESSMENT

<b>Delivery method</b>	In person.	
<b>USE OF INFORMATION AND COMMUNICATION TECHNOLOGIES</b>		
<b>TEACHING ORGANIZATION</b>	<b>Activity</b>	<b>Semester Workload</b>
	Lectures	39 (13 week x 3 hours)
	Laboratory exercises.	26 (13 Lb. Ex. x 2 hours)
	Report of laboratory exercises	21
	Preparation for written exam	39 (13 Lect. x 2 hours)
	<b>Total Course (25 workload hours per credit unit)</b>	<b>125 (5 ECTS)</b>
<b>STUDENT EVALUATION</b>	<p>I. Written exam (80 %) of graded difficulty including:</p> <ul style="list-style-type: none"> <li>- Multiple choice questions</li> <li>- Short questions</li> <li>- Questions of crisis and development</li> </ul> <p>II. Laboratory exercises (20%):</p> <ul style="list-style-type: none"> <li>- Participation and performance during the laboratory exercise</li> <li>- Written report of laboratory results</li> </ul> <p>Therefore: the total grade is obtained as a sum of the above two individual evaluations.</p>	

#### 5. SUGGESTED BIBLIOGRAPHY

*-Suggested Bibliography :*

- Molecular Techniques in Food Biology: Safety, Biotechnology, Authenticity and Traceability Aly Farag El Sheikha (Editor), Robert E. Levin (Editor), Jianping Xu (Editor)
- The Use of Molecular Biology Techniques in Food Traceability. M. Espiñeira, F.J.Santaclara

*-Related Scientific Journals:*

Journal of Microbiology, Biotechnology and Food Sciences  
 Food Science and Technology  
 Molecular Nutrition and Food Research  
 Molecular Nutrition and Food Technology  
 Molecular gastronomy  
 Food Chemistry